



University  
of Dundee

# From GWAS to Functional Studies in South Indian Population

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**How do I leverage our GWAS results, a  
goldmine of wealth ? ???**

*An experimental approach...*

**“Low levels of HDL Cholesterol is a key Metabolic difference in Indian population compared to European population”**

*Reinforced from WP2 data*

*CETP*(Cholesteryl Ester Transfer Protein ) , *LIPC* (Hepatic lipase ) and *LPL* (Lipoprotein lipase ) **WP2 RESULTS**



Analyze and find the rare variants from T2D in Indian population



Validate by Functional studies and finally prove the variants as a Functional variants or otherwise.

# FUNCTIONAL CHARACTERIZATION OF GENETIC VARIANTS

(Both for EOT2D variants and HDL pathway project )

## WORKFLOW

Choosing the rare variants



Creation of variants using Site Directed Mutagenesis (SDM)



Transformation (using top10 *E.coli* cells)



Confirmation of mutagenesis by Colony PCR and Sequencing



Isolation of Plasmids



Confirmation of plasmids using sequencing



Functional Studies based on the nature of gene product

# GWAS TO FUNCTIONAL STUDIES

To characterize genes coding for Transcription Factors:

Reporter assay

To access the transcription activity.

Western blot

To access the protein expressions.

EMSA

To access the DNA binding activity.

Subcellular localization

To access the localization of protein.

To characterize genes coding for channel proteins:

Western blot

To access the protein expressions.

Surface expression

To check the trafficking of channel.

Thallium flux assay

To check the channel response to known inhibitors/pharmacological agents.

To characterize genes coding for enzymes:

Western blot

To access the protein expressions.

Kinetic assay

To access/measure the half life of an enzyme.

Activity assay

To access/measure the enzyme activity.

# Molecular cloning of Genetic variants-SITE DIRECTED MUTAGENESIS (SDM)

Primer designing (<https://www.agilent.com/store/primerDesignProgram.jsp>)

1. It is recommended that you clear this form prior to loading each new sequence:

Clear Input

2. Select QuikChange® mutagenesis kit that you are using:

QuikChange® II

[Help in choosing a mutagenesis kit](#)

3. Find your DNA sequence by pressing

Load a file from your hard-drive:

No file selected

Browse\_

Or, paste plain text or FASTA-formatted DNA sequence in the box below:

4. Load it.

Upload Now

- or -

Upload Translated

or specify a DNA region to translate from

to

Upload Translated Region

Finally, obtain your designed primer sequences.

Design Primers

Site Directed Mutagenesis using *Quick-change II Site-Directed Mutagenesis Kit*



Transformation (*E.coli* top10 competent cells)



Colony PCR



Cycle sequencing



Confirmation of variants



Plasmids isolation



Plasmids will be ready



# TRANSACTIVATION ASSAY

Cells will be plated



Incubate at CO2 incubator at 37°C for 18-20hrs



Confluency (70%-80%)



Cells will be transfected



Cells will be Incubate at CO2 incubator at 37°C for 20-24hrs



Cell Lysate will be prepared by using Passive Lysis Buffer(PLB)



Reporter Assay

Wild type-pcDNA 3.1-WT

Target DNA -pGL3-target DNA

Internal control-pRL -SV40

Variant- pcDNA3.1-V1

# WESTERN BLOTTING

Cells will be plated



Confluency (70%-80%)



Cells will be transfected (Wild type & variant)



Incubate at CO<sub>2</sub> incubator at 37°C for 24-36hrs



Cell Lysate will be prepared by using RIPA Lysis Buffer in cold conditions

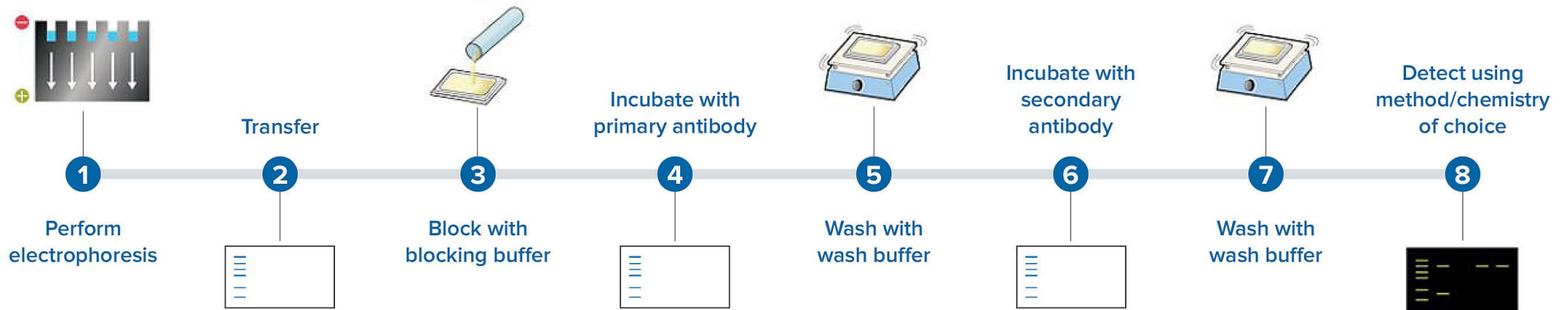


SDS-PAGE

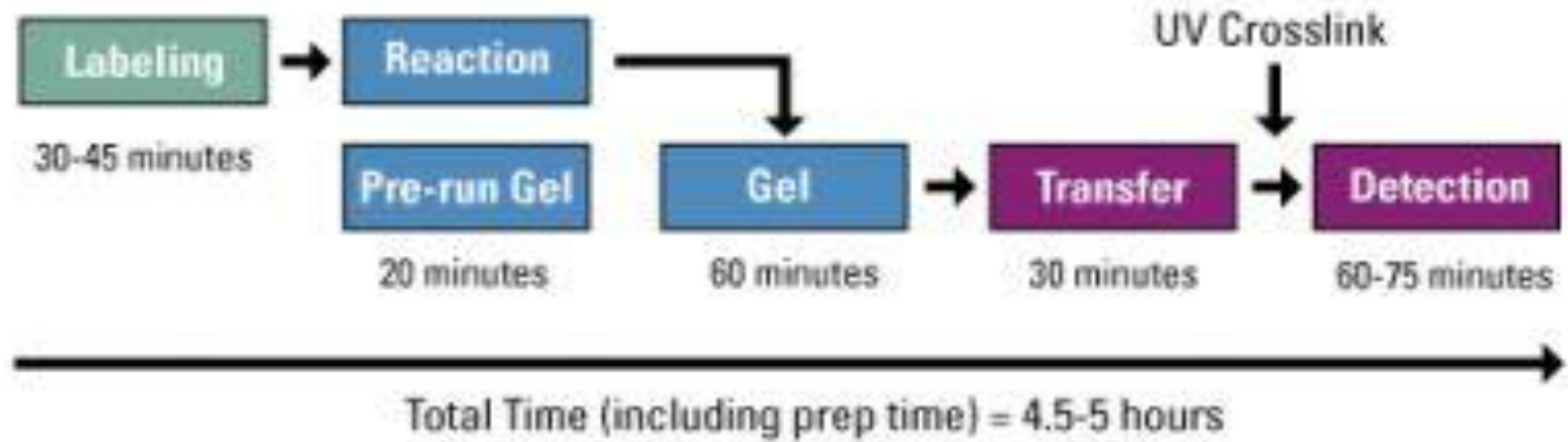


# WESTERN BLOTTING OR IMMUNOBLOTTING

- Tissue preparation      RIPA Lysis buffer
- Gel electrophoresis      SDS-PAGE (Polyacrylamide Gel Electrophoresis)
- Transfer      To the membrane ( PVDF or Nitrocellulose membranes)
- Blocking      Bovine serum albumin
- Detection      Primary and secondary antibody
- Analysis



# Electrophoretic Mobility Shift Assay (EMSA)



**Electrophoretic Mobility Shift Assay: Analyzing Protein - Nucleic Acid Interactions**  
Carolina Alves and Celso Cunha

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